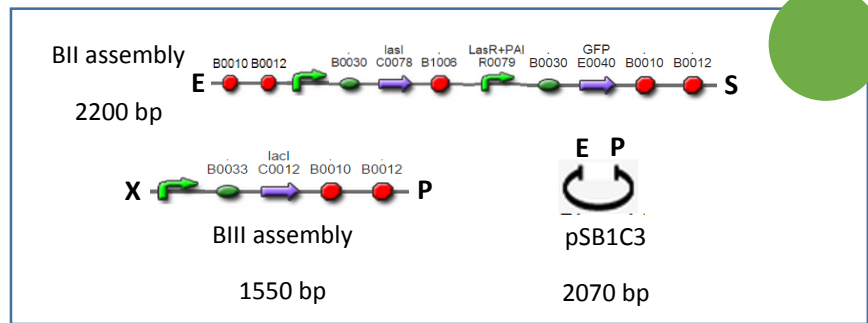


Assembly:



1st Day

EXSP Digestion (see **Enzymatic Digestion Protocol**)

| Parts | ng/ul | Volume to 2,5 ug (ul) | Buffer x10 (ul) | EcoRI (ul) | XbaI (ul) | SpeI (ul) | PstI (ul) | H ₂ O to 50ul (ul) |
|---------------|-------|-----------------------|-----------------|------------|-----------|-----------|-----------|-------------------------------|
| BII assembly | 196,6 | 13,0 | 5 | 1 | - | 1 | - | 30 |
| BIII assembly | 156,9 | 16,0 | 5 | - | 1 | - | 1 | 27 |
| pSB1C3 | 107,3 | 24,3 | 5 | 1 | - | - | 1 | 20,7 |

Repeat this digestion only if you run out of stock

2nd Day

Gel Purification

- See **Kit Wizard SV gel and PCR clean up Promega Protocol**
- Quantify digestion products

| Parts | ng/ul | 260/280 |
|--------------------|-------|---------|
| BII assembly (ES) | 25,5 | 1,89 |
| BIII assembly (XP) | 18,8 | 2,05 |
| pSB1C3 (EP) | 24,3 | 2,83 |

Obs: 260/280 is a quality parameter that tells you if your sample is contaminated with proteins. The greater it is compared to 1 the less contaminants you have.

Ligation (see **Ligation Protocol**)

| | | |
|---|------|------|
| Linear Plasmid 50 ng | 2 ul | |
| Insert : Plasmid 3:1 (BII) ; 3:1 (BIII) | BII | BIII |
| | 6 ul | 6 ul |
| 10x T4 DNA Buffer | 2 ul | |
| T4 DNA ligase 1u | 1 ul | |
| H ₂ O to 20 ul | 3 ul | |

Obs: To determinate the amount of DNA necessary we used the following equation

$$\text{Insert ng} = \text{plasmid ng} \times \frac{\text{insert bp}}{\text{plasmid bp}} \times \text{insert:plasmid ratio}$$

- Incubate overnight at 37°C.
- Prepare and sterilize in the autoclave tubes with 6 ml of liquid LB medium
- Prepare glycerol 40%

3rd Day

Transformation (see **Transformation Protocol in *Escherichia coli* DH5-α**)

Organism: *E. coli* DH5-α

Selection: Cloranphenicol

4th Day

- Inoculate 3 – 4 colonies in a 6 ml LB with the same antibiotic used in the transformation protocol.
- Incubate overnight at 275rpm/37°C.

5th Day

Miniprep

- Prepare **glycerol stock** of the clones (500ul glycerol 40% + 500ul inoculum).
- Extract plasmidial DNA (see **Alkaline Lyses or PureLink Invitrogen Protocol**)
- Run a preliminary electrophoresis gel.
- Quantify DNA samples.

Assembly Confirmation

- EP Digestion (see **Enzymatic Digestion Protocol**)

| Assembly | Volume to 300 ng (ul) | Buffer x10 (ul) | EcoRI (ul) | PstI (ul) | H ₂ O to 10ul (ul) |
|----------|-----------------------|-----------------|------------|-----------|-------------------------------|
| CII – 1 | 3 | 1 | 0,5 | 0,5 | 5 |
| CII – 2 | 3 | 1 | 0,5 | 0,5 | 5 |
| CII – 3 | 3 | 1 | 0,5 | 0,5 | 5 |

- Incubate for 2 hours at 37°C.
- Prepare samples for DNA sequencing.
- Run an electrophoresis analysis of the EP digestion



| Size expected | Size in gel |
|---------------|-------------|
| 3750 bp | ~ 3700 bp |